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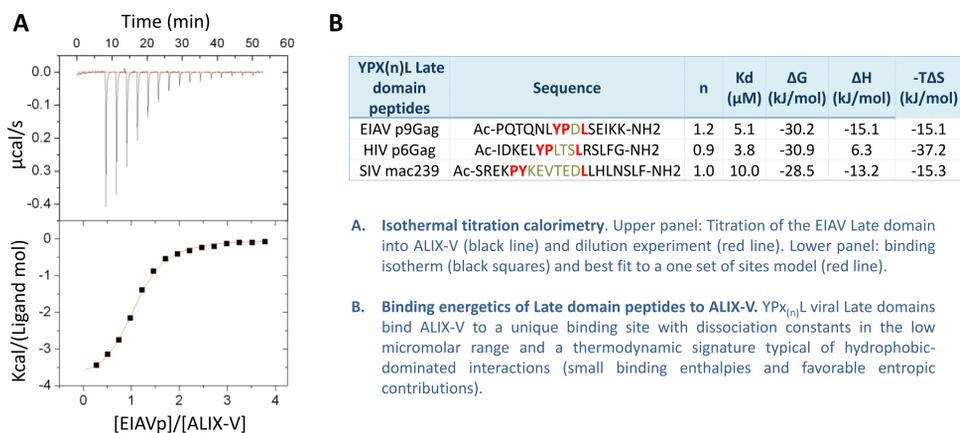
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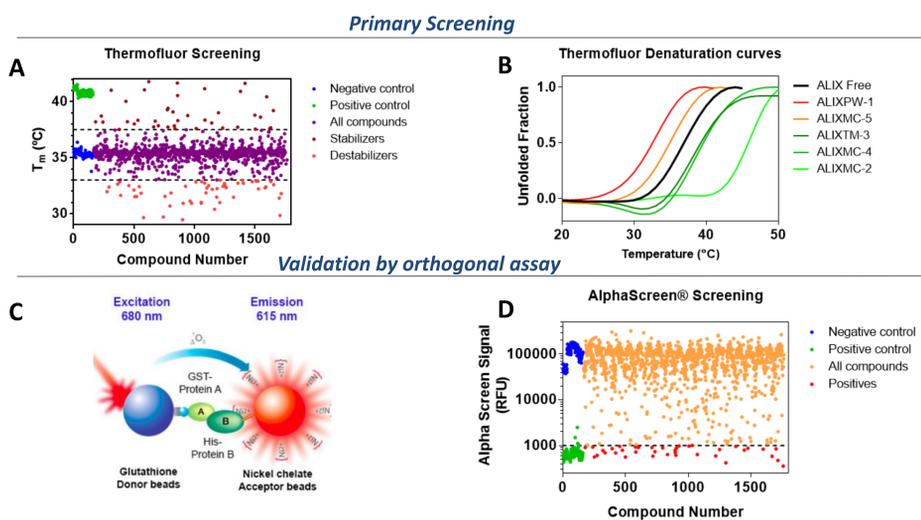
ABSTRACT

Most membrane-enveloped viruses (including retroviruses and RNA viruses) bud from infected cells by recruiting the host ESCRT machinery through the direct interaction of short and conserved viral sequences called Late domains with some cellular factors. Small-molecule inhibitors of these interactions efficiently block the egress of a number of viruses and are interesting candidates as novel broad-spectrum antivirals. LYP(x)nL Late domains bind to the V domain of the adaptor protein ALIX (ALIX-V), which is a fairly unexplored domain. We present here a thermodynamic study of the molecular determinants of ALIX-V peptide recognition using Isothermal Titration Calorimetry and MicroScale Thermophoresis together with a phage-display analysis of the binding preferences in ALIX-V peptide ligands. Also, the druggability of ALIX-V has been assessed by the high-throughput screening of large compound libraries in search for small-molecule inhibitors of ALIX-V/Late domain interactions. The screening of different compound and drug-repurposing libraries using virtual screening methodologies combined with miniaturized Thermofluor and AlphaScreen® assays has allowed the identification of a set of compounds that bind ALIX-V with IC50 values in the low micromolar range. Bimolecular Complementation Assays validated some of these compounds as ALIX-V/Late domain interactions inhibitors in a cellular context. In order to further explore the druggability of ALIX-V in a richer chemical space we have developed miniaturized HTRF® and AlphaScreen® assays for the screening of the large library of actinomycetes and fungi extracts of the Fundación MEDINA. A pilot assay of a subset of extracts has allowed the identification of one novel compound with IC50 in the low micromolar range. This work provides new insights of the interaction of ALIX-V with the viral proteins and paves the way towards the development of novel broad-spectrum antivirals.

1. The binding of YPX_(n)L viral Late domains to ALIX-V is weak and entropically driven, suggesting a highly hydrophobic interaction

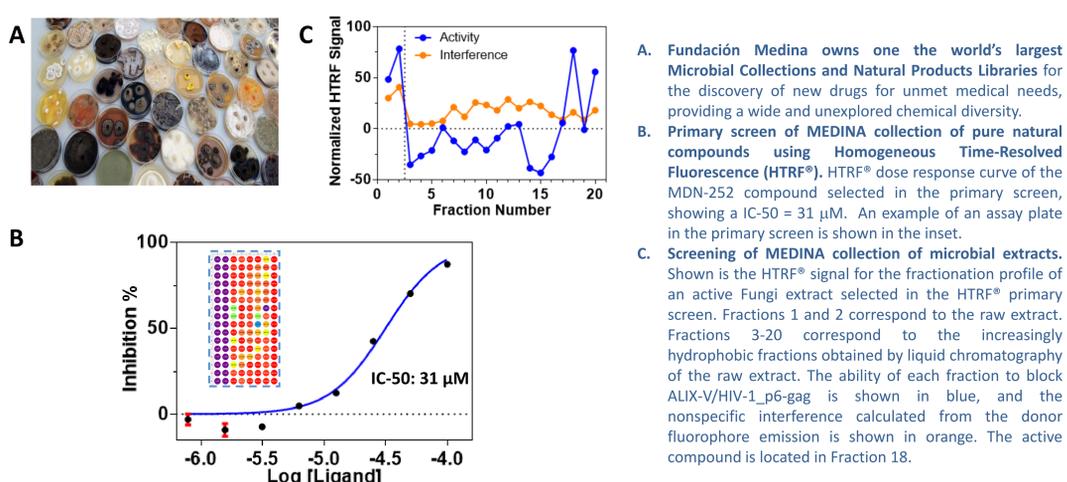


2. ALIX-V and p6-Gag Late domain interactions can be blocked by small molecules

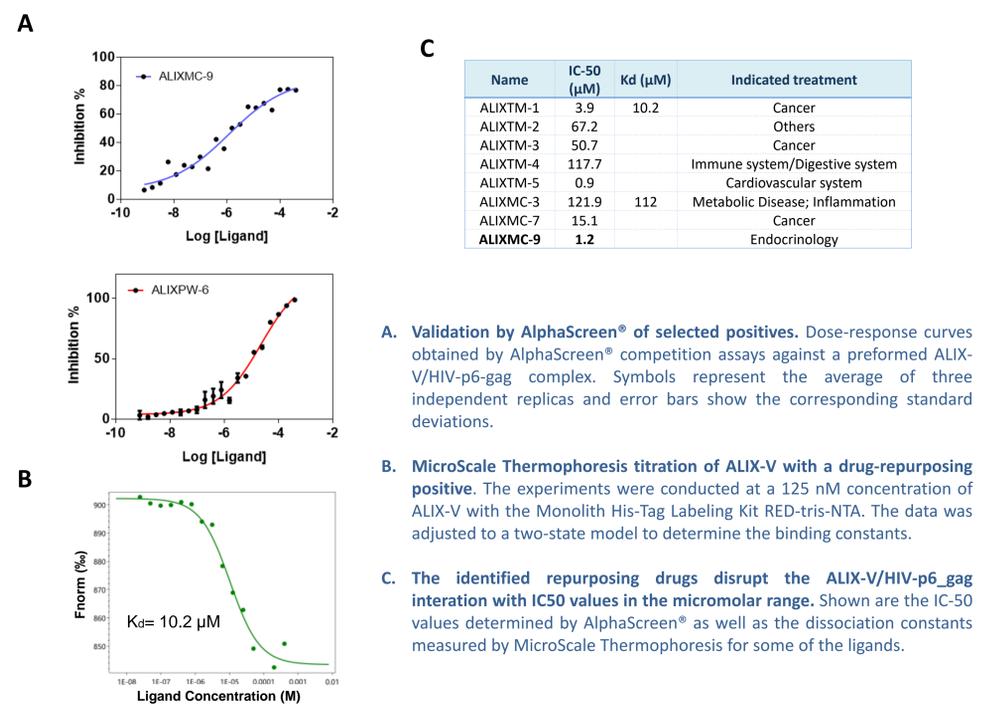


- A. Primary high-throughput screening of drug repurposing libraries using differential scanning fluorimetry (DSC-Thermofluor)** to identify small-molecule binders of ALIX-V. Compounds capable of changing the denaturation temperature of ALIX-V to higher than 37.5° C or lower than 33° C were selected for further study.
- B. Normalized DSF profiles for ALIX-V.** Shown are the normalized traces of the free ALIX-V domain (black) and some stabilizing (green) and destabilizing (red/orange) ligands.
- C. AlphaScreen® was used as a secondary orthogonal assay for validation.** The His-tagged ALIX-V protein interacts with a Nickel chelate bead as the GST-tagged p6-Gag protein (HIV) interacts with a Glutathione bead. The interaction between His-tagged ALIX-V and GST-tagged HIV_p6-Gag protein provides a readable signal. The presence of compounds blocking this interaction leads to loss of luminescence signal.
- D. Orthogonal high-throughput screening of drug repurposing libraries by AlphaScreen®** to identify small-molecule able to disrupt the ALIX-V/HIV-p6-gag interaction. Compounds that lowered the RFU below 1000 units were selected. A 40% coincidence between the two screening campaigns was obtained.

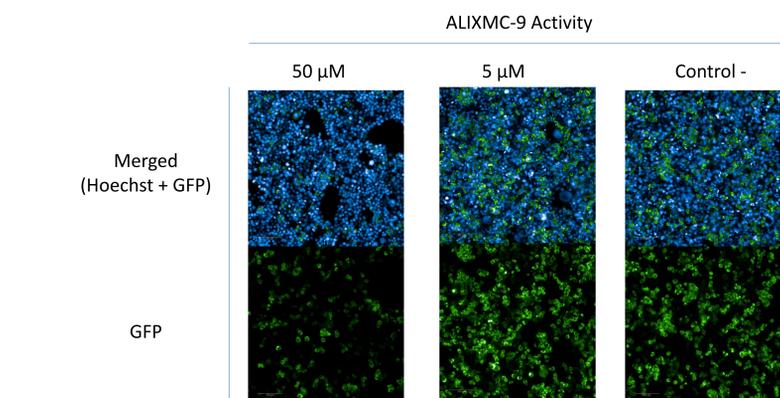
3. A screening of the Fundación MEDINA collection of actinomycetes and fungi extracts has allowed the identification of natural products with the ability to disrupt ALIX-V/HIV_p6-gag interaction.



4. A small set of repurposing drugs can disrupt the ALIX-V/HIV-1-p6-gag with IC50 values in the micromolar range



5. Small-molecule inhibitors can block the interaction between the full-length ALIX-V and HIV_p6-gag proteins in a cellular context



Cellular Images of the Bimolecular Fluorescence Complementation (BiFC) Cellular Assays. Green Fluorescence Protein (GFP) signal is produced upon interaction between the ALIX and HIV_p6 proteins. Some of the identified ALIX ligands were found to block the interaction between the full-length proteins without significant toxicity. The figure contains the results after ALIXMC-9 treatment at 50 and 5 μM.

Conclusions

- The thermodynamic characterization show new insights that are key to understand ALIX-V interactions and to approach the design of optimized, high-affinity ligands.
- The ALIX-V binding interface is druggable. Drug repurposing molecules and natural compounds with the ability to block the interaction between ALIX-V and viral late domains in vitro have been identified, which show good potential as novel broad-spectrum antivirals.
- Our screening methodology can readily provide new compounds with the ability to disrupt the host-virus proteins interaction in a cellular context, validating the target for antiviral discovery.

Funding: BIO2012-39922-CO2, BIO2016-78746-C2-1-R; Spanish Ministry of Science and Innovation. CV20-19149; Universities, Research and Technology Department, Junta de Andalucía.

